

ab119211– InstantBlue® Coomassie Protein Stain (ISB1L)

Specially formulated for ultra-fast, sensitive, and safe detection of your proteins.
For research use only - not intended for diagnostic use.

This product is manufactured by Expedeon, an Abcam company, and was previously called and was previously called InstantBlue Protein Stain (and also Optiblot Blue). ISB1L is the same as the 1000 ml size.

For overview, typical data and additional information please visit:

<https://www.abcam.com/ab119211>

(use www.abcam.cn/ab119211 for China, or www.abcam.co.jp/ab119211 for Japan)

Storage and Stability: Upon receipt store at 4°C. Once used, the staining solution should be discarded and cannot be reused.

Materials Supplied:

Item	Quantity	Storage temperature
InstantBlue® Protein Stain	1000 mL	4°C

ΔNote: InstantBlue® Protein Stain contains Coomassie dye, ethanol, phosphoric acid and solubilizing agents in water. **(Caution: Phosphoric acid is a corrosive liquid.)**

Reagent Preparation: Provided as ready-to-use solution and should not be diluted.

Technical Hints:

- Multiple washes prior to staining with InstantBlue® are NOT required or recommended.
- An alcohol/acetic acid fixing step prior to staining with InstantBlue® is NOT required or recommended.
- A destaining step post staining is NOT required or recommended with InstantBlue®.

Staining Protocol:

Mix the InstantBlue® solution immediately before use by gently inverting the bottle a few times (do not shake the bottle to mix the solution).

Standard Protocol:

1. After electrophoresis remove the gel from the tank and transfer directly into the InstantBlue® staining solution. Be sure that the gel moves freely in stain to facilitate diffusion. Typically, ~20 mL is needed to cover the gel.
2. Coloured protein bands will start to develop immediately, and a suitable intensity is typically achieved after 15 minutes incubation at room temperature with gentle shaking.
3. Photograph your gel when the required intensity has been achieved. Gels can be kept in staining solution but ensure that the gel remains covered with liquid. Alternatively, the gel can be stored in ultrapure water after staining for 1 hour in InstantBlue®.

Protocol for gel drying:

1. Ensure that the gel has been staining for at least 1 hour.

Δ Note: Although protein bands will be visible after a few minutes of incubation in stain, the staining process is typically fully completed after 1h incubation. Depending on the type of gel you are using, longer incubation may be necessary. Further processing of the gel prior to completion of the staining process may result in protein destaining and reduced sensitivity. If this occurs, simply restain the gel by incubating overnight in InstantBlue®.

2. Submerge the gel in approximately 100 mL ultrapure water at ~70°C (heat for 30s to 60s in a microwave oven). Incubate for at least 1 hour while gently rocking. Optionally, adsorbent paper or paper towel can be added. Gels can be incubated overnight in ultrapure water.
3. Incubate the gel in a 'gel drying solution' (e.g. 4% glycerol, 20% ethanol in water) for 2 minutes. Incubation of any Coomassie-stained gel in an alcohol solution will eventually result in destaining of the bands, so avoid incubation for longer than 5 minutes.
4. The gel is now ready for drying between wetted cellophane membranes.

Protocol for destaining protein bands for MS analysis:

1. Excise the protein band of interest and transfer to a clean sample tube.
2. Add 1 ml of 30% ethanol or 30% acetone or 30% acetic acid

ΔNote: Acetic acid may result in acetylation of the N-terminus.

3. Incubate for 20 min (incubate at 60°C – 70°C to increase the rate of destaining).
4. Decant supernatant and repeat steps 2 & 3 at least 3 times, or until gel is clear.

Technical Support

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